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The Effectiveness of Super Ovulation and Multiple Pregnancies in Sprague Dawley Rat using *Morus alba* Linn. Fruit

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ABSTRACT

The white mulberry, Morus alba L. has been used by several Asian societies for the treatment of infertility. However, there is no evidence that products of the plant can influence ovarian health. Thus, the aim of this study was to determine the effect of the Morus alba fruit extract on the ovarian function of nonpregnant and pregnant rats. The study showed that in rats Morus alba fruit extract stimulates follicular stimulating (FSH) and luteinizing hormones (LH), estrogen, and progesterone productions that peaked at 8 days of treatment. The effect was dose-dependent with hormone production increasing with increase in dose of the extract. The Graafian follicles were fully matured, and the number increased with increase in dose of the extract. The uterus of pregnant rats contained several embryos that gave birth to the full-term offspring without abortion or embryonic abnormalities. In conclusion, the Morus alba fruit extract has potential to be developed into a compound for the treatment of female infertility.

Keywords: Ovulation induction, Multiple pregnancies, Hormone levels, Natural products, Rat model

INTRODUCTION

The Chinese has a long history of traditional use mulberry as herb medicine [1]. They have used the fruit for treating many ailments from premature greying of hair to food poisoning and hypertension [2,3]. The *Morus alba*, also known as white mulberry (Figure 1), is a fast-growing, small to medium-sized flowering tree belonging to the family Moraceae. The tree is widely distributed and cultivated in most Eastern Asian countries [4] and certain temperate regions of the world, including Kurdistan region-Northern Iraq. The tree is a good source of food for humans and animals and the leaves are used to feed silkworms [1]. The mulberry juice or mulberry pomace is used as a diet supplement by virtue of its isocaloric and isonitrogenous properties similar to corn grain and cottonseed meals [5]. Although clinical trials on the use of the *Morus alba* contract has been conducted, the full potential of the tree as a source of therapeutic compounds has yet to be fully realized [6].



Figure 1(a) Morus alba fruit on the tree



Figure 1(b) *Morus alba* fruit in dried form

The various biological and pharmacological activities attributed to the mulberry fruit, leaves and roots include antioxidation [7-10], macrophage activation [11], antimicrobial infection [12-14], antivirus infection [15], antiparasitic [16], anti-inflammation [17-19], cytotoxicity and anticancer [17-22], anti-diabetes [23-25], anti-ulcer [26], anti-hyperlipidemia [27], anti-atherosclerosis [28-31], anti-obesity [32,33], anti-aging [34], anti-hypertension [35], cardioprotection [36], cognitive enhancement, neuroprotection [37], and hepatoprotection [38,39]. Other pharmacological properties of *Morus alba* include anti-platelet [40], anxiolytic, anti-asthmatic [41], antidepressant, and immunomodulatory activities [6].

Mulberry leaves have flavonoids as major constituents with polyphenols in lesser amounts [42]. The root bark of the *Morus alba* contains flavonoids, alkaloids, and stilbenoids [43,44] and the mulberry fruit is rich in linoleic, palmitic, oleic, and stearic acids [45].

The treatment of choice for anovulation in infertile women has been clomiphene citrate (CC), which is a relatively safe and effective oral drug. However, this drug is known to have side effects such as psychological disorder [46], hot flashes and ovarian cysts [47], and is implicated a cause of visual disturbance [48].

In Iran, *Morus alba* is listed as one of the plant in Iranian traditional medicine used in the treatment of infertility in men [49]. On the contrary, in India the juice of the *Morus alba* leaves is taken orally as a contraceptive. In the rat diabetic model, it was shown that oral treatment with cooked pulverized mulberry leaves increased the number of germinal, Leydig, and Sertoli cells, diameter of seminiferous tubules, testosterone level, and testis weight [50]. Currently, there is no reported evidence on the effect of products of the mulberry plant on the female reproductive system. Thus, our aim is to determine the effect of the *Morus alba* fruit extract on the reproductive activities of female rats.

MATERIALS AND METHODS

Fruit Extract Preparation

The fruit of *Morus alba* L. (Moraceae) collected from uncultivated fields in the vicinity of Sulaimani city, Kurdistan region, Northern Iraq during fruiting season of May to June 2016. The harvested fruit authenticated in the Department of Field Crop, College of Agricultural Sciences, University of Sulaimani with voucher number UNIVSUL/AGRI/

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FC/0011. The fruit was sun-dried to brown in color and grounded into coarse powder. Exactly 250 g of *Morus alba* fruit powder was subjected to the maceration extraction method [51]) using 500 mL ethanol. The ethanol filtrate was evaporated for 48 hours using the Buchi vacuum rotary evaporator (Sigma-Aldrich) to obtain a semi-solid mass, which was kept in an air-tight glass container and stored at 4°C.

Experimental Design

Thirty-two 7 to 8 weeks old virgin female Sprague-Dawley rats weighing between 150 g and 200 g and were obtained from the Animal House of Department of Clinic and Internal Medicine, College of Veterinary Medicine, University of Sulaimani, Iraq. The rats were kept in polystyrene cage and acclimatized for 5 days under room temperature of $25 \pm 1^{\circ}$ C with a 10 to14-hour light-dark cycle and normal rat diet and water were provided ad libitum. Rats were grouped into four (A to D, n=8 each). Group A served as control negative and received normal rat diet and water, whereas groups B, C, and D rats received 300 mg/kg, 600 mg/kg, and 900 mg/kg of freshly prepared semisolid *Morus alba* fruit extract in 1 mL of distilled water daily. The extract was administered orally using force feeding needle once daily for 14 days.

On day 7 of treatment, 2 fertile male rats were introduced to each cage for 24 hours to allow mating. Two rats from each group were euthanized on day 7 to obtain the ovaries and another 2 on day 15 to obtain the developing embryos. The embryos were macroscopically examined. The rest of the pregnant rats were allowed to carry to term to deliver their pups. Euthanasia was conducted humanely using chloroform inhalation anesthetic in a tightly closed glass chamber and the ovaries and embryos collected. The study was approved by the Animal Care and Use Committee, College of Veterinary Medicine, University of Sulaimani (UNIVSUL/ACUC/2014-0099).

Histopathological Examination of the Ovaries

The Ovaries were fixed in 10% of formalin for at least 48 hours before placing in plastic cassettes and dehydrated using an automated tissue processor (Leica, ASP300, Germany). The tissues were embedded in paraffin wax, and the blocks sectioned using a semi-automated microtome (Leica, RM2155, Germany). Subsequently, the tissue sections mounted on the glass slides were deparaffinized and rehydrated. The sections were stained with Harris's hematoxylin and eosin (H&E) [52]. Finally, tissue sections mounted with cover slips using Distrene-Plasticizer Xylene (DPX) were examined using a light microscope image analyzer (Olympus BX51TF Japan).

Hormonal Measurement

Approximately 3 mL of blood was collected from each rat via cardiac puncture, while under Di-ethyl ether anesthesia, on days 0, 8 and 15 of the experiment in EDTA. Plasma was obtained for FSH, LH, estrogen and progesterone measurement using the ELISA method [53].

Statistical Analysis

The results of this experiment expressed as mean \pm SD and analyzed statistically using SPSS version 20.0 (SPSS Inc., Chicago, USA). Significance level was determined at α =0.05.

RESULTS

The effect of treatment with *Morus alba* fruit extract on the rat plasma LH, FSH, and estrogen was peak and most significant (p<0.05) after 8 days (Figures 4-7). The level of these hormones decreased again after 15 days. The effect of the *Morus alba* fruit extract was dose-dependent with the concentration of hormones positively correlated with concentration of extract used in the treatment. The plasma LH and FSH peaked at 1.56 ± 0.01 , 0.53 ± 0.08 , mIU/mL, respectively while estrogen at 101.03 ± 0.6 pg/mL at Day 8 after treatment with 900 mg/kg of extract. The effect of treatment with *Morus alba* fruit extract on plasma progesterone increased significantly (p<0.05) both with treatment concentration and period of treatment. The highest plasma progesterone was 99.67 \pm 0.25 pg/mL after treatment with 900 mg/kg extract at Day 15. The extract appears to have a delayed effect on the plasma progesterone (Table 1).

Table 1(a) Mean values of FSH, LH, estrogen and progesterone levels following administration of *Morus alba* extract on Day 0

Group	FSH (mIU/ml)	LH (mIU/ml)	Estrogen (pg/ml)	Progesterone (pg/ml)
A (Control Negative)	0.18 ± 0.44	0.88 ± 0.11	56.07 ± 0.3	18.20 ± 0.10
B (Low Dose)	0.15 ± 0.31	0.75 ± 0.55	52.31 ± 0.1	19.32 ± 0.73
C (Mid Dose)	0.16 ± 0.17	0.82 ± 0.15	49.03 ± 0.3	17.16 ± 0.51
D (High Dose)	0.19 ± 0.30	0.79 ± 0.01	54.39 ± 0.9	20.31 ± 0.90

Values are given as mean \pm SEM for each group; There is no significant difference (p<0.05) in all treatment groups compared to Group A; Statistical level of significance was determined by one-way ANOVA

Table 1(b) Mean values of FSH, LH, estrogen and progesterone levels following administration of Morus alba extract on Day 8

Group	FSH (mIU/ml)	LH (mIU/ml)	Estrogen (mIU/ml)	Progesterone (mIU/ml)
A (Control Negative)	0.19 ± 0.09	0.86 ± 0.07	54.06 ± 0.35	19.20 ± 1.2
B (Low Dose)	$0.28 \pm 0.05*$	$1.10 \pm 0.15*$	77.36 ± 1.2*	22.4 ± 0.8
C (Mid Dose)	$0.41 \pm 0.12*$	$1.34 \pm 0.11*$	$89.07 \pm 0.3*$	25.52 ± 0.5
D (High Dose)	$0.53 \pm 0.20*$	$1.56 \pm 0.01*$	$101.03 \pm 0.6*$	29. 71 ± 0.07*

Values are given as mean \pm SEM for each group; * indicates a significant difference (p<0.05) in treated groups compared to group A (control group); Statistical level of significance was determined by one-way ANOVA

Table 1(c) Mean values of FSH, LH	f. estrogen and progesterone leve	els following administration of M	forus alba extract on Day 15
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Group	FSH (mIU/ml)	LH (mIU/ml)	Estrogen (mIU/ml)	Progesterone (mIU/ml)
A (Control Negative)	0.17 ± 0.01	0.85 ± 0.91	56.07 ± 0.9	18.21±1.07
B (Low Dose)	0.20 ± 0.05	0.90 ± 0.15	60.36 ± 1.2	38.21± 0.01*
C (Mid Dose)	0.24 ± 0.12	0.94 ± 0.11	64.07 ± 0.13	$63.44 \pm 0.08*$
D (High Dose)	$0.29\pm0.30*$	$0.99 \pm 0.01*$	70.03± 0.04*	$99.67 \pm 0.25*$

Values are given as mean \pm SEM for each group; * indicates a significant difference (p<0.05) in treated groups compared to Group A; Statistical level of significance was determined by one-way ANOVA

The histopathological examination showed that non-treated immature virgin rats had follicles at intermediate stage of development (Figure 2A). However, the ovaries of rats treated with 300 mg/kg *Morus alba* fruit extract had both fully matured and secondary Graafian follicles. The matured follicles contained fluid filled cavities (Figure 2B). The Graafian follicles was highest in number in rats treated with 900 mg/kg extract suggesting the effect directly dose-dependent (Figures 2C and 2D).

The ovaries and uterine horns of non-treated virgin rats were normal in size and gross morphology (Figure 3A). Premated rats sacrificed on day 8 of treatment showed enlarged ovaries with clear, brilliant, good blood supply, healthy uterus and uterine horns (Figure 3B). After mating on Day 15 of treatment, pregnant rats showed multiple embryos (Figure 3C). By Day 21 of treatment with *Morus alba* fruit extract the embryos were still healthy and well-nourished (Figure 3D).

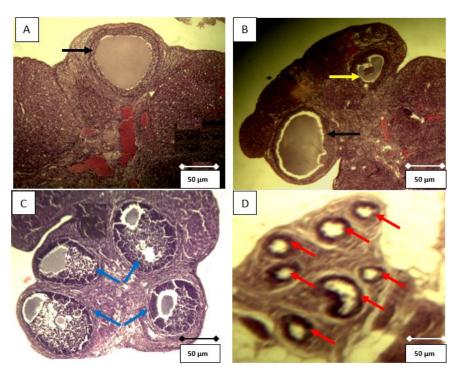


Figure 2 The effect of treatment with *Morus alba* on oocyte maturation and superovulation in Sprague-Dawley Rats. A) Control negative group without treatment. Black arrow: follicle at intermediate stage of development. B) 300 mg/kg *Morus alba* treatment group. Black arrow: Mature follicle. Yellow arrow: follicle at intermediate stage of development. C) 600 mg/kg *Morus alba* treatment group. Blue arrow: Multiple Graafian follicles. D) 900 mg/kg *Morus alba* treatment group. Red arrows: Multiple follicles at various stages of development

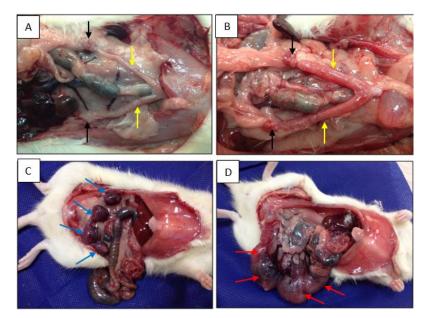


Figure 3 Viscera of Sprague-Dawley rats treated with 900 mg/kg *Morus alba*. A) Normal control negative group without treatment showing normal-sized ovaries and oviducts. B) Virgin rat treated for 1 week showing enlarged ovaries and good blood supply in both ovaries. Black arrows: Ovaries. Yellow arrows: Oviducts. C) Pregnant rats treated for 2 weeks. Blue arrows: multiple embryos. D) Pregnant rats treated for 3 weeks. Red arrows: multiple developed embryos

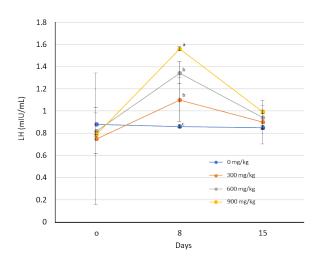


Figure 4 Plasma luteinizing hormone level in rats treated with *Morus alba* extract. Each point represents mean with standard deviations. For each time period, means with different letters are significantly different at p<0.05

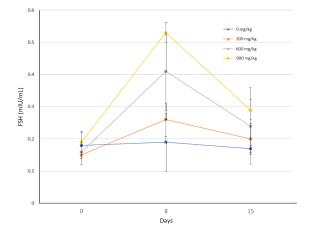


Figure 5 Plasma follicle-stimulating hormone level in rats treated with *Morus alba* extract. Each point represents mean with standard deviations. For each time period, means with different letters are significantly different at p<0.05

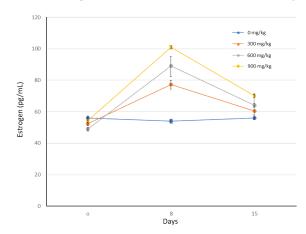


Figure 6 Plasma estrogen level in rats treated with *Morus alba* extract. Each point represents mean with standard deviations. For each time period, means with different letters are significantly different at p<0.05

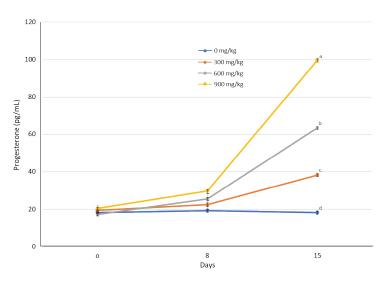


Figure 7 Plasma progesterone level in rats treated with *Morus alba* extract. Each point represents mean with standard deviations. For each time period, means with different letters are significantly different at p<0.05

DISCUSSION

The first choice of treatment in the management of infertility in normally estrogenized, an ovulatory woman is clomiphene citrate (CC). The drug improves reproductive hormonal milieu by competing with estrogen at the hypothalamic level. Clomiphene citrate causes an increase in gonadotropin-releasing hormone (GnRH) from the hypothalamus resulting in increased secretion of FSH and LH from the anterior pituitary gland [54].

Among the Kurdish, fresh and dried mulberry fruit are used a means to balance sex hormones, treat infertility, and a support for embryo health. Although the fruit is purported to be of health benefit, its effective has not been verified through controlled investigations. Currently, it is not known as to effect of the plant extract on the female reproductive hormones. In this study, the rat was used as a model to determine the efficaciousness of the M. alba fruit extract in the maintenance of pregnancy. Our study showed that consumption of the mulberry fruit extract did not cause any ill-effects on pregnant rats.

The pregnant rat is an excellent model for the investigation on female reproductive cycle by virtue that rat has a short estrous cycle lasting approximately four days. The proestrus and estrus phases of the estrous cycle are characterized by increases in plasma FSH and LH levels that are associated with folliculogenesis and ovulation. The FSH and LH stimulate theca cells during folliculogenesis to stimulate production and release of estradiol [55]. Steroidogenesis in preovulatory follicles occurs via LH receptors on the theca cells and FSH receptors, possibly also involving LH receptors, on the granulosa cells. This shows that FSH and LH play important roles in follicle development. In the present study, treatment with *Morus alba* fruit extract had increased production of FSH and LH after 8 days. The level of these hormones decreased again after 15 days of treatment.

FSH stimulates production of estrogens by the granulosa cells of the ovarian follicles and corpus luteum. The increase in production of ovarian FSH in rats as the result of treatment with *Morus alba* fruit extract had enhanced the secretion of estrogen hormone from the granulose cells. However, the effect of FSH on significant progesterone production as the result of *Morus alba* fruit extract treatment occurs later than the effect of estrogen production; most probably approximately one week later. It has also been suggested that increased FSH production stimulate follicular growth, and this can occur without the contribution of LH [56].

Histopathologically, *Morus alba* fruit extract treatment did not affect the size or morphology of Graafian follicles. However, the *Morus alba* fruit extract had increased the number of fully matured follicles and this effect is dosedependent, that is, greater number of follicles with treatment with higher doses of extract. This phenomenon is expected to result in superovulation and multiple pregnancies. This result indicates that both increasing levels of FSH and LH hormones played a critical role in both superovulation and multiple pregnancies. The increase in follicle number as the result of treatment with extract was accompanied enlargement of ovary, increase in blood supply, activated and engorged uterus, which are conditions conducive for multiple pregnancy. The maintenance of the superovulated ovary is supported by increase in estrogen and progesterone production.

CONCLUSION

This study shows that *Morus alba* fruit extract can potentially be used to stimulate ovulation that will facilitate multiple pregnancies while contributing to the maintenance of embryo health. In conclusion, *Morus alba* fruit extract may be used to increase sex hormone production in females, improve ovulation that could lead to superovulation and multiple pregnancies. The extract has potential to be developed as a safe alternative source in the treatment of infertility in females.

DECLARATIONS

Conflict of Interest

Authors declared that there is no conflict of interest. The authors alone are responsible for the content of this manuscript.

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REFERENCES

- [1] Younus, Ishrat, et al. "A review of ethnobotany, phytochemistry, antiviral and cytotoxic/anticancer potential of *Morus alba* linn." *International Journal of Advanced Research and Review*, Vol. 1, No. 2, 2016, pp. 84-96.
- [2] Huo, Yongkang. "Mulberry cultivation and utilization in China." *Mulberry for Animal Production. FAO Animal Production and Health Paper*, Vol. 147, 2002, pp. 11-43.
- [3] Wei, Hua, et al. "Review of bioactive compounds from root barks of Morus plants (Sang-Bai-Pi) and their pharmacological effects." *Cogent Chemistry*, Vol. 2, No. 1, 2016, p. 1212320.
- [4] Devi, Bandna, et al. "Morus alba Linn: A phytopharmacological review." International Journal of Pharmacy and Pharmaceutical Sciences, Vol. 5, No. 2, 2013, pp. 14-18.
- [5] Zhou, Zhenming, et al. "Effect of ensiled mulberry leaves and sun-dried mulberry fruit pomace on finishing steer growth performance, blood biochemical parameters, and carcass characteristics." *PLoS ONE*, Vol. 9, No. 1, 2014, p. e85406.
- [6] Chan, Eric Wei-Chiang, Lye Phui-Yan, and Wong Siu-Kuin. "Phytochemistry, pharmacology, and clinical trials of *Morus alba*." *Chinese Journal of Natural Medicines*, Vol. 14, No. 1, 2016, pp. 17-30.
- [7] Arfan, Muhammad, et al. "Antioxidant activity of mulberry fruit extracts." *International Journal of Molecular Sciences*, Vol. 13, No. 2, 2012, pp. 2472-80.
- [8] Imran, Mohammad, et al. "Chemical composition and antioxidant activity of certain Morus species." Journal of Zhejiang University-Science B, Vol. 11, No. 12, 2010, pp. 973-80.
- [9] Liang, Linghong, et al. "Chemical composition, nutritional value, and antioxidant activities of eight mulberry cultivars from China." *Pharmacognosy Magazine*, Vol. 8, No. 31, 2012, p. 215.
- [10] Wang, Wei, et al. "In vitro antioxidant and antimicrobial activity of extracts from Morus alba L. leaves, stems and fruits." The American Journal of Chinese Medicine, Vol. 40, No. 02, 2012, pp. 349-56.
- [11] Kim, Seon Beom, et al. "Macrophage activating activity of pyrrole alkaloids from *Morus alba* fruits." *Journal of Ethnopharmacology*, Vol. 145, No. 1, 2013, pp. 393-96.
- [12] De Oliveira, Alisson Macário, et al. "Evaluation of toxicity and antimicrobial activity of an ethanolic extract from leaves of Morus alba L. (Moraceae)." Evidence-Based Complementary and Alternative Medicine, No. 2015, 2015.
- [13] Sheikhlar, Atefeh, et al. "White mulberry (*Morus alba*) foliage methanolic extract can alleviate Aeromonas hydrophila infection in African Catfish (*Clarias gariepinus*)." *The Scientific World Journal*, No. 2014, 2014.
- [14] Thabti, Ines, et al. "Phenols, flavonoids, and antioxidant and antibacterial activity of leaves and stem bark of Morus species." *International Journal of Food Properties*, Vol. 17, No. 4, 2014, pp. 842-54.

- [15] Du, Jiang, et al. "Antiviral flavonoids from the root bark of Morus alba L." Phytochemistry, Vol. 62, No. 8, 2003, pp. 1235-38.
- [16] Liang, Jing-Han, et al. "Identification and effect of two flavonoids from root bark of Morus alba against Ichthyophthirius multifiliis in grass carp." Journal of Agricultural and Food Chemistry, Vol. 63, No. 5, 2015, pp. 1452-59.
- [17] Eo, Hyun Ji, et al. "Anti-inflammatory and anti-cancer activity of mulberry (Morus alba L.) root bark." BMC Complementary and Alternative Medicine, Vol. 14, No. 1, 2014, p. 200.
- [18] Jing, Wanghui, et al. "Mori Cortex regulates P-glycoprotein in Caco-2 cells and colons from rats with experimental colitis via direct and gut microbiota-mediated mechanisms." *RSC Advances*, Vol. 7, No. 5, 2017, pp. 2594-2605.
- [19] Park, Geun-Soo, Jeong-Keun Kim, and Jae-Hong Kim. "Anti-inflammatory action of ethanolic extract of Ramulus mori on the BLT2-linked cascade." *BMB Reports*, Vol. 49, No. 4, 2016, p. 232.
- [20] Naowaratwattana, Wanlaya, Wanchai De-Eknamkul, and Elvira Gonzalez De Mejia. "Phenolic-containing organic extracts of mulberry (*Morus alba* L.) leaves inhibit HepG2 hepatoma cells through G2/M phase arrest, induction of apoptosis, and inhibition of topoisomerase IIα activity." *Journal of Medicinal Food*, Vol. 13, No. 5, 2010, pp. 1045-56.
- [21] Qin, Jing, et al. "New cytotoxic and anti-inflammatory compounds isolated from *Morus alba* L." *Natural Product Research*, Vol. 29, No. 18, 2015, pp. 1711-18.
- [22] Radojković, Marija, et al. "Biological activities and chemical composition of Morus leaves extracts obtained by maceration and supercritical fluid extraction." *The Journal of Supercritical Fluids*, Vol. 117, 2016, pp. 50-58.
- [23] Mudra, Mitchell, et al. "Influence of mulberry leaf extract on the blood glucose and breath hydrogen response to ingestion of 75 g sucrose by type 2 diabetic and control subjects." *Diabetes Care*, Vol. 30, No. 5, 2007, pp. 1272-74.
- [24] Wang, Hsiu-Ju, and Been-Huang Chiang. "Anti-diabetic effect of a traditional Chinese medicine formula." Food & Function, Vol. 3, No. 11, 2012, pp. 1161-69.
- [25] Xu, Lingyuan, et al. "Anti-diabetic effect mediated by *Ramulus mori* polysaccharides." *Carbohydrate Polymers*, Vol. 117, 2015, pp. 63-69.
- [26] Abdulla, Mahmood Ameen, et al. "Evaluation of the anti-ulcer activities of *Morus alba* extracts in experimen-tally-induced gastric ulcer in rats." *Biomedical Research*, Vol. 20, No. 1, 2009.
- [27] El-Beshbishy, Hesham A., et al. "Hypolipidemic and antioxidant effects of *Morus alba* L. (Egyptian mulberry) root bark fractions supplementation in cholesterol-fed rats." *Life Sciences*, Vol. 78, No. 23, 2006, pp. 2724-33.
- [28] Al-Shehabi, Tuqa Saleh, Rabah Iratni, and Ali H. Eid. "Anti-atherosclerotic plants which modulate the phenotype of vascular smooth muscle cells." *Phytomedicine*, Vol. 23, No. 11, 2016, pp. 1068-81.
- [29] Chan, Kuei-Chuan, et al. "Mulberry leaf extract inhibits the development of atherosclerosis in cholesterol-fed rabbits and in cultured aortic vascular smooth muscle cells." *Journal of Agricultural and Food Chemistry*, Vol. 61, No. 11, 2013, pp. 2780-88.
- [30] Chao, Py., et al. "Inhibitive effects of mulberry leaf-related extracts on cell adhesion and inflammatory response in human aortic endothelial cells." *Evidence-Based Complementary and Alternative Medicine*, Vol. 2013, 2013.
- [31] Enkhmaa, Byambaa, et al. "Mulberry (*Morus alba* L.) leaves and their major flavonol quercetin 3-(6-malonylglucoside) attenuate atherosclerotic lesion development in LDL receptor-deficient mice." *The Journal of Nutrition*, Vol. 135, No. 4, 2005, pp. 729-34.
- [32] Choi, Ji Won, et al. "Structural analysis and anti-obesity effect of a pectic polysaccharide isolated from Korean mulberry fruit Oddi (*Morus alba* L.)." Carbohydrate Polymers, Vol. 146, 2016, pp. 187-96.
- [33] Lim, Hyun Hwa, et al. "Combined treatment of mulberry leaf and fruit extract ameliorates obesity-related inflammation and oxidative stress in high fat diet-induced obese mice." *Journal of Medicinal Food*, Vol. 16, No. 8, 2013, pp. 673-80.
- [34] Zheng, Shanqing, et al. "Mulberry leaf polyphenols delay aging and regulate fat metabolism via the germline signaling pathway in *Caenorhabditis elegans*." *Age*, Vol. 36, No. 6, 2014, p. 9719.
- [35] Yang, Nae-Cherng, Kun-Yan Jhou, and Chin-Yin Tseng. "Antihypertensive effect of mulberry leaf aqueous extract containing γ-aminobutyric acid in spontaneously hypertensive rats." Food Chemistry, Vol. 132, No. 4, 2012, pp. 1796-1801.
- [36] Lee, Jung-Jin, et al. "Morusinol extracted from *Morus alba* inhibits arterial thrombosis and modulates platelet activation for the treatment of cardiovascular disease." *Journal of Atherosclerosis and Thrombosis*, Vol. 19, No. 6, 2012, pp. 516-22.
- [37] Kang, Tong Ho, et al. "Neuroprotective effects of the cyanidin-3-O-β-d-glucopyranoside isolated from mulberry fruit against cerebral ischemia." *Neuroscience Letters*, Vol. 391, No. 3, 2006, pp. 122-26.

- [38] Hsu, Li-Sung, et al. "Mulberry water extracts (MWEs) ameliorated carbon tetrachloride-induced liver damages in rat." Food and Chemical Toxicology, Vol. 50, No. 9, 2012, pp. 3086-93.
- [39] Sun, Xufeng, et al. "Effects of quercetin derivatives from mulberry leaves: Improved gene expression related hepatic lipid and glucose metabolism in short-term high-fat fed mice." *Nutrition Research and Practice*, Vol. 9, No. 2, 2015, pp. 137-43.
- [40] Kim, Dong-Seon, et al. "Antiplatelet activity of *Morus alba* leaves extract, mediated via inhibiting granule secretion and blocking the phosphorylation of extracellular-signal-regulated kinase and akt." *Evidence-Based Complementary and Alternative Medicine*, No. 2014, 2014.
- [41] Jung, Hyo Won, et al. "Effect of kuwanon G isolated from the root bark of *Morus alba* on ovalbumin □ induced allergic response in a mouse model of asthma." *Phytotherapy Research*, Vol. 28, No. 11, 2014, pp. 1713-19.
- [42] Zou, Yuxiao, et al. "Phenolics and antioxidant activity of mulberry leaves depend on cultivar and harvest month in Southern China." *International Journal of Molecular Sciences*, Vol. 13, No. 12, 2012, pp. 16544-53.
- [43] Jung, Jae-Woo, et al. "Three new isoprenylated flavonoids from the root bark of *Morus alba*." *Molecules*, Vol. 21, No. 9, 2016, p. 1112.
- [44] Wei, Hua, et al. "Review of bioactive compounds from root barks of Morus plants (Sang-Bai-Pi) and their pharmacological effects." Cogent Chemistry, Vol. 2, No. 1, 2016, p. 1212320.
- [45] Sánchez-Salcedo, Eva M., et al. "Fatty acids composition of Spanish black (Morus nigra L.) and white (Morus alba L.) mulberries." Food Chemistry, Vol. 190, 2016, pp. 566-71.
- [46] Choi, So-Hyun, et al. "Psychological side-effects of clomiphene citrate and human menopausal gonadotrophin." Journal of Psychosomatic Obstetrics & Gynecology, Vol. 26, No. 2, 2005, pp. 93-100.
- [47] Kistner, Robert W., John L. Lewis, and Gillen J. Steiner. "Effects of clomiphene citrate on endometrial hyperplasia in the premenopausal female." *Cancer*, Vol. 19, No. 1, 1966, pp. 115-26.
- [48] Viola, Maria I., David Meyer, and Thinus Kruger. "Association between clomiphene citrate and visual disturbances with special emphasis on central retinal vein occlusion: A review." *Gynecologic and Obstetric Investigation*, Vol. 71, No. 2, 2011, pp. 73-76.
- [49] Nejatbakhsh, Fatemeh, et al. "Recommended foods for male infertility in Iranian traditional medicine." Iranian Journal of Reproductive Medicine, Vol. 10, No. 6, 2012, p. 511.
- [50] Muchtaromah, Bayyinatul, et al. "Effect of mulberry (Morus alba L.) leaves infusion on the reproductive status of chronic diabetic model." Journal of Applied Environmental and Biological Sciences, Vol. 5, No. 5, 2015, pp. 14-18.
- [51] Trusheva, Boryana, Dorina Trunkova, and Vassya Bankova. "Different extraction methods of biologically active components from propolis: A preliminary study." *Chemistry Central Journal*, Vol. 1, No. 1, 2007, p. 13.
- [52] Luna, Lee G., editor. Manual of Histologic Staining Methods of the Armed Forces Institute of Pathology. Blakiston Division, McGraw-Hill, 1968.
- [53] Matsui, Hisanori, et al. "Peripheral administration of metastin induces marked gonadotropin release and ovulation in the rat." *Biochemical and Biophysical Research Communications*, Vol. 320, No. 2, 2004, pp. 383-88.
- [54] Tulandi, Togas, et al. "Congenital malformations among 911 newborns conceived after infertility treatment with letrozole or clomiphene citrate." *Fertility and Sterility*, Vol. 85, No. 6, 2006, pp. 1761-65.
- [55] Tabak, Joël, et al. "Variations in the response of pituitary lactotrophs to oxytocin during the rat estrous cycle." *Endocrinology*, Vol. 151, No. 4, 2010, pp. 1806-13.
- [56] Macklon, Nick S., et al. "The science behind 25 years of ovarian stimulation for *in vitro* fertilization." *Endocrine Reviews*, Vol. 27, No. 2, 2006, pp. 170-207.